

Single or Daily Application of Topical Curcumin Prevents Ultraviolet B-Induced Apoptosis in Mice

by Indah Puspasari

Submission date: 09-Jan-2023 11:41AM (UTC+0700)

Submission ID: 1990026661

File name: 1673239284_13_Single_or_Daily_Application_of_Topical_Curcumin_Prevents.pdf (900.22K)

Word count: 4429

Character count: 27127

Article

Single or Daily Application of Topical Curcumin Prevents Ultraviolet B-Induced Apoptosis in Mice

Khairuddin Djawad ¹, Ilham Jaya Patellongi ^{2,*}, Upik Anderiani Miskad ³, Muhammad Nasrum Massi ⁴, Irawan Yusuf ² and Muhammad Faruk ⁵

¹ Department of Dermatology and Venereology, Faculty of Medicine, Hasanuddin University, Makassar 90245, Indonesia

² Department of Physiology, Faculty of Medicine, Hasanuddin University, Makassar 90245, Indonesia

³ Department of Pathological Anatomy, Faculty of Medicine, Hasanuddin University, Makassar 90245, Indonesia

⁴ Department of Microbiology, Faculty of Medicine, Hasanuddin University, Makassar 90245, Indonesia

⁵ Department of Surgery, Faculty of Medicine, Hasanuddin University, Makassar 90245, Indonesia

* Correspondence: ilhamjaya@unhas.ac.id

Abstract: Curcumin is a natural ingredient with antioxidant effects, widely studied as a treatment for various types of cancer. However, its effects on ultraviolet radiation have not been fully explored. The effects of single or daily application of 0.1–100 μM curcumin on cell apoptosis in ultraviolet B (UVB)-induced mice were tested using an experimental double-blind posttest design with a control group and two research models: a single application of curcumin before a single UVB exposure and daily application of curcumin for 7 days before a single UVB exposure on the seventh day. Apoptotic cells were counted using a tunnel system kit. The number of apoptotic cells under a single or daily application of curcumin for 7 days was significantly lower than that of the UVB controls ($p \leq 0.05$). The number of apoptotic cells decreased with the increasing concentration of curcumin, and the maximum effect was observed at 100 μM. Daily application of topical curcumin was superior in preventing apoptosis (mean apoptotic cell count of 14.86 ± 1.68) compared with a single application (17.46 ± 0.60 ; $p = 0.011$). Topical curcumin can act as a potential photoprotective agent in preventing cutaneous malignancies due to UVB radiation. Further studies are warranted, especially in humans.

Keywords: curcumin; radiation; apoptosis



Citation: Djawad, K.; Patellongi, I.J.; Miskad, U.A.; Massi, M.N.; Yusuf, I.; Faruk, M. Single or Daily Application of Topical Curcumin Prevents Ultraviolet B-Induced Apoptosis in Mice. *Molecules* **2023**, *28*, 371. <https://doi.org/10.3390/molecules28010371>

Academic Editor: Federica Belluti

Received: 24 November 2022

Revised: 17 December 2022

Accepted: 22 December 2022

Published: 2 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Curcumin is a polyphenolic compound extracted from turmeric (*Curcuma longa*) and is a yellowish pigment that contains various metabolites, such as dihydrocurcumin, tetrahydrocurcumin, hexahydrocurcumin, octahydrocurcumin, curcumin glucuronide, and curcumin sulfate. This compound has been used as a chemotherapeutic or chemopreventive agent and an alternative therapy for various diseases [1]. Curcumin possesses antioxidant, anti-inflammatory, immunomodulatory, hepatoprotective, anti-ischemic, nephroprotective, antimicrobial, hypoglycemic, and antirheumatic properties [2]. It also has anticarcinogenic effects related to its molecular activity of inducing the apoptosis of cancerous cells but interestingly sparing healthy cells. This compound has been used in the therapy of breast, pancreatic, and colorectal cancers, and multiple myeloma [1,3].

Topical curcumin at a dose of 30 μM can induce apoptosis in several tumor cells, thereby reducing cell cycle progression and preventing the growth of cancer cells [4]. The apoptotic effect of curcumin depends on the concentration and duration of its administration. In terms of morphology, the apoptotic cells exhibit several characteristic changes compared with normal cells, such as shrinkage, pyknosis, and plasma membrane blebbing, leading to the formation of apoptotic bodies [1,5]. In contrast to its effect on cancerous cells, curcumin inhibits the apoptosis of keratinocyte cells in ultraviolet-induced cells [6].

However, the research on its influence on ultraviolet B (UVB) radiation is still limited. Previous studies used keratinocytes as the main target of UVB exposure to explore photodamage and apoptosis [7] and found that curcumin can prevent the accumulation of the abnormal cells that cause skin cancer [7–9]. The application of curcumin prior to UVB exposure in mice suppressed erythema and apoptosis by inhibiting p53 and interleukin-6 (IL-6) expression and increasing anti-inflammatory IL-10 expression in the pretreated skin compared with those in the controls [8]. Information regarding the anti-apoptotic effect of topical curcumin is limited, especially in UVB-induced cell apoptosis.

This study aimed to explore the preventive effect of topical curcumin on UVB-induced mice by histopathologically examining the number of apoptotic cells using two different study designs. The first group was applied with topical curcumin in four concentrations once, followed by a single UVB exposure. The second group was applied with topical curcumin in four concentrations once daily for 7 consecutive days, followed by a single UVB exposure. Biopsy was then performed to determine the number of apoptotic cells. The optimal number of applications and concentration were also determined for the potential use of topical curcumin as a photoprotective agent.

2. Results

The weight of all mice was homogenous and did not differ significantly across all test groups ($p = 0.76$). Thus, the difference in the effect of all topical curcumin concentrations for all the treatment groups could be further analyzed. ANOVA showed an apoptosis-inhibitory effect in both research models ($p \leq 0.05$). Post hoc analysis revealed that the number of apoptotic cells in the non-UVB-exposed group was significantly different from that in the UVB and acetone controls ($p \leq 0.05$). All groups that received the four concentrations of topical curcumin in either single or daily applications with subsequent UVB exposure showed significantly lower apoptotic cell counts compared with the UVB control group ($p \leq 0.05$). Significant differences in apoptotic cell count were observed among the curcumin groups with various concentrations ($p \leq 0.05$), except for the 1 μM group ($p > 0.478$). A similar inhibitory effect against UVB-induced apoptosis was noted in the second experimental model (Figure 1).

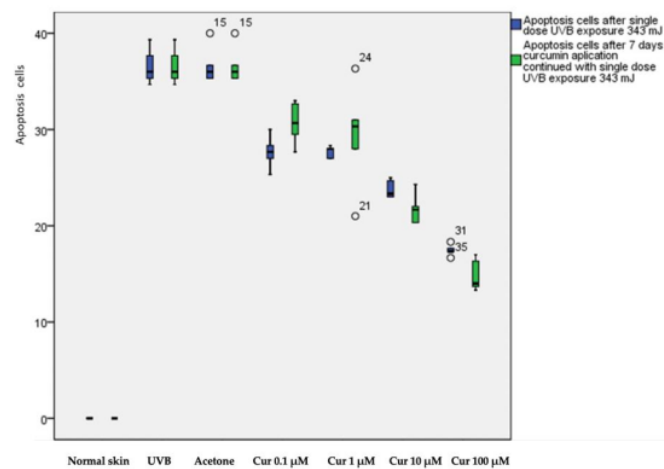


Figure 1. Induction of apoptosis in UVB-irradiated mice. Pretreatment with topical curcumin before UVB irradiation prevented apoptosis in both experimental models.

Table 1 exhibits the difference in apoptotic cell count between the single application of topical curcumin of various concentrations with a subsequent single UVB exposure and the daily application of topical curcumin for 7 days with a subsequent single UVB exposure.

The *t*-test showed a statistically significant difference in apoptotic cell count between the one-time and seven-time curcumin applications among all concentration groups ($p < 0.05$) except the 1 μM group. All curcumin concentrations in a single application resulted in a decrease in the apoptotic cell count. The number of cells was similar among the various concentrations except for 100 μM . When curcumin was applied daily for 7 consecutive days, the number of apoptotic cells decreased significantly under treatment concentrations of 10 μM and 100 μM with a mean of 21.73 ± 1.64 and 14.86 ± 1.68 , respectively ($p = 0.041$, 0.011). At low concentrations of 0.1 μM and 1 μM , the single application of curcumin (mean number of apoptotic cells: 27.86 ± 1.78 and 27.76 ± 0.96 , respectively) was superior to the daily application (35.07 ± 2.23 and 29.33 ± 5.57 , respectively). However, only the results for 0.1 μM were statistically significant ($p = 0.042$).

Table 1. Comparison of apoptotic cell count between one-time and seven-time curcumin application before single 343-mJ UVB exposure.

Concentration	Curcumin Number of Applications	Number of Apoptotic Cells		<i>p</i> -Value
		Mean \pm SD	Mean Difference	
0.1 μM	1	27.86 ± 1.78		
	7	30.70 ± 2.23	−2.84	0.042 *
1 μM	1	27.76 ± 0.96		
	7	29.33 ± 5.57	−1.57	0.478
10 μM	1	23.80 ± 0.96		
	7	21.73 ± 1.64	2.07	0.041 *
100 μM	1	17.46 ± 0.60		
	7	14.86 ± 1.68	2.60	0.011 *

Note: * Significant; SD, Standard deviation.

Histopathological examination showed that regardless of the number of applications, an increase in curcumin concentration led to a decrease in cell apoptosis compared with the positive control. This finding revealed the inhibitory effect of topical curcumin, which was most prominent at the 100 μM concentration after the seven-time application (Figure 2A,B).

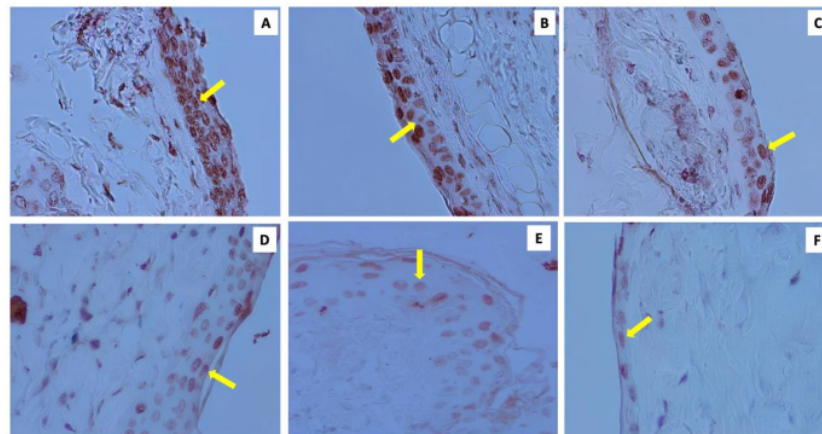


Figure 2. Apoptotic cells, indicated by brown staining in the nuclei (arrow), at 400 \times magnification under an Olympus Type CX-31 microscope, in the UVB-only group with majority apoptosis cells (A), acetone (B), 0.1 μM concentration (C), 1 μM concentration (D), 10 μM concentration (E), and 100 μM concentration with the least amount of cell apoptosis (F).

Based on histopathological examination, with an increase in curcumin concentration, both in single and daily application, the results showed a decrease in cell apoptosis com-

pared to the positive control. This showed the inhibitory effect of topical curcumin, most prominently in the 100 μ M concentration after the seven-time application (Figure 2A,B).

3. Discussion

Cell apoptosis or programmed cell death serves as an integral part of cellular homeostasis. An increase in the abnormal viability of cells influenced by endogenous or exogenous factors can lead to the development of various diseases, most prominently malignancies and autoimmune diseases. The protein family BCL-2 is responsible for regulating cellular apoptosis balance through pro-apoptotic and pro-survival members and thus serves as a basis for potential therapeutic developments for various diseases [10]. Cell apoptosis occurs through two pathways; extrinsic or cytoplasmic pathway triggered by Fas death receptors and intrinsic or mitochondrial pathways triggered by the release of cytochrome C from mitochondria. One of the most important proteins in this pathway is BCL-2. The overexpression of this protein leads to cell accumulation in the G0 phase [11]. Although apoptosis is a defense mechanism to prevent cell mutations that can lead to malignancies, the prevention of cell damage or repairing of cell damage through normal cell mechanism in cases of mild radiation is preferred [12,13].

The usefulness of curcumin has long been explored, and the compound is associated with modulation on various pathways. Special interest has been directed to the autophagy or degradation of superfluous or dysfunctional components within cells, in which curcumin inhibits the formation of reactive oxygen species (ROS) and acts as an antioxidant agent [14,15]. The topical use of curcumin is ideal compared with systemic administration; even though the compound has a high safety profile, it is poorly absorbed systemically and has a rapid elimination [16]. Curcumin can either induce or inhibit cellular apoptosis in various malignant cells and concentrations, such as in human melanoma cells (30–60 mM for 4 h), human leukemia (HL) 60 cells (10–40 mM for 16–24 h), AK-5 tumor cells (10 mM for 16 h), and MCF-7 breast cancer cells (25 mM for 24 h). Apoptosis inhibition was observed in dexamethane-induced apoptosis in rat thymocytes and chemotherapy-induced apoptosis in breast cancer cells [17]. An evaluation of the efficacy of curcumin for treating infantile hemangioma endothelial cells (HemECs) found that its 10 μ M concentration exhibited a high inhibition activity for the proliferation capability of HemECs; as proven by positive annexin-V-FITC staining, caspase-3 activation, and the cleavage of poly(ADP-ribose) polymerase (PARP) in the treated cells, curcumin achieved low-micromolar IC₅₀ (half maximal inhibitory concentration) and induced apoptosis in HemECs through the downregulation of myeloid cell leukemia-1 (MCL-1) and hypoxia-inducible factor 1 α [18]. Furthermore, curcumin is useful in treating psoriasis by reducing epidermal thickness, erythema, pruritus, and burning and pain sensations [19].

Minimal differences in apoptotic cell count were observed among the three curcumin concentrations of 0.1, 1, and 10 μ M in single application with subsequent UVB radiation. However, a drastic decrease was found when the concentration was increased to 100 μ M. In addition, the highest concentration (100 μ M) yielded the lowest number of apoptotic cells after daily use for 7 consecutive days, providing the best result in this study. Although an increase in application and concentration seemed to result in an overall low apoptotic cell count, the single application of the low concentrations of 0.1 and 1 μ M was superior compared with their daily application. We have yet to establish the reason behind this phenomenon. However, we hypothesized that this phenomenon may be related to the biphasic effect of curcumin on the oxidation of post-prandial chylomicrons and its biphasic hormetic response on proteasome activity and heat-shock protein synthesis in human keratinocytes [20,21]. Furthermore, we postulated that the cumulative effect of antioxidants during the daily application of topical curcumin enhances the inhibitory effect on cell damage and apoptosis, as reported by Zhou et al. [22]. Another study regarding the pre-treatment oral curcumin during a seven-day period prevented cyclophosphamide-induced lung injury in rats through the suppression of oxidative stress, therefore reducing the number of cell apoptosis [23].

¹⁸ The protective effect of curcumin in this research was supported by other experimental studies. Li et al. [16] reported that pretreatment with curcumin effectively inhibited photo-damage in mice and human keratinocyte (HaCaT) cells using a UVB dose of 540 mJ/cm² (3 MED) for 3 consecutive days. Different results were obtained by Park et al. [24], who also pretreated HaCaT cells with curcumin before exposing them to a subapoptotic dose of UVB (100 mJ/cm²) to induce apoptosis. The difference in results may be due to the differences in UVB doses. The pretreatment of curcumin prior to UVB radiation can reduce the number of ROS by acting as a scavenger for most ROS [14,16]. Although the production of ROS after UVB radiation is a natural and protective response toward UVB radiation, the number of ROS released should be controlled or reduced because an excessive amount of oxidative stress can damage various cellular compounds, such as nucleic acids, proteins, and lipids. It can lead to a ³² G-C mutation that acts as a precursor for photoaging and cutaneous malignancies [25]. Reactive oxygen species also play an important role in UVA damage. A study evaluating the efficacy of pretreatment with topical curcumin before UVA radiation found that this compound significantly decreased the level of NF- κ B, a protein that plays an integral role in the inflammation pathway [26]. Furthermore, curcumin has moderate inhibition properties toward proteins MMP-1 and MMP-3, which are expressed after UVA and UVB radiation, resulting in the restoration of collagen metabolism, wound healing, and regulating disorders [26,27]. Recently, Li et al. [16] and Barball ²⁸ et al. [19] found that curcumin prevents apoptosis by inhibiting the ³⁶ production of ROS through the modulation of the Nrf2 and NF- κ B signaling pathways. Nrf2 is a key transcription factor in oxidative stress that can influence the activation of antioxidant response by various cyto⁶ protective and antioxidant enzyme genes [6,28–30].

In the current study, we observed that the application of topical curcumin gave a protective effect against ultraviolet exposure-induced apoptosis in mice. However, this study has limitations because it did not examine the markers of oxidative stress (reactive oxygen species, hydrogen peroxide, and malondialdehyde) and inflammation (cyclooxygenase-2, interleukin, prostaglandin E2, tumor necrosis factor-alpha, and nitric oxide) that may have an effect on the process of apoptosis.

4. Methods

4.1. Study Design and Subject

¹ The study was a double-blind experimental posttest design with a control group, conducted at the Hasanuddin University Animal Laboratory, Makassar, Indonesia. Health ³⁴ male Swiss albino mice aged 6–9 weeks and weighing 20–30 g were maintained under a temperature of 28 °C \pm 2 °C and humidity of 50% \pm 10% for a minimum of 1 week. All mice were shaved on the back regularly during the research.

The topical preparation was curcumin purchased ¹⁰ from Sigma-Aldrich, Inc. (St Louis, MO, USA). It was diluted in acetone at concentrations of 0.1 μ M, 1 μ M, 10 μ M ⁶ and 100 μ M. The UVB lamp source was 10 FS-40-T12 fluorescent sun lamps with a spectrum of 280–340 nm and a peak emission of 314 nm. The UVB lights were calibrated with a FLUX radiometer.

4.2. Study Protocol

Two experimental models were used: a single application of various curcumin concentrations before a single UVB exposure and daily application of various curcumin doses for 7 days before a single UVB exposure on the seventh day. The mice were randomly allocated to 11 groups, each containing five mice. Group 1 received no treatment, group 2 received only UVB irradiation, and group 3 received acetone and UV ¹⁰ irradiation. Groups 4–7 received topical curcumin applications with concentrations of 0.1 μ M, 1 μ M, 10 μ M, and 100 μ M, respectively, with a dose of 2 μ L/cm² on the back for 20 min before exposure to 343 mJ of UVB. Groups 8–11 received topical curcumin applications at similar concentrations once daily for 7 days and were irradiated with 343 mJ of UVB 20 min after the last topical curcumin application on the seventh day. Skin biopsy for histopathological

examination was performed 24 h after UVB exposure, and the obtained samples were assessed under 400× magnification.

4.3. Cell Apoptosis Activity Examination

The apoptotic cells among the squamous epithelial cells in all epidermis layers were counted histopathologically using the Apo-BrdU-IHC Kit TUNEL System (Biovision®, Milpitas, CA, USA), following the manufacturer protocol. After being deparaffinized and rehydrated, the tissue samples were permeabilized with proteinase K (30 µg/mL) for 20 min at room temperature and then removed from endogenous peroxidase activity using 3% hydrogen peroxide for 5 min. After being rinsed with phosphate-buffered saline (PBS), the samples were incubated with 5× reaction buffer for 10 min, added to DNA labeling solution, and incubated at 37 °C for 1.5 h. The reaction was stopped by immersing the tissue in PBS twice within 15 min. An anti-BrdU-Biotin solution was added to each slide and incubated for 1.5 h. The samples were washed with PBS and incubated with streptavidin-HRP for 30 min at room temperature before being rinsed again. After rinsing, the tissues were incubated with diaminobenzidine for approximately 10 min until a change to a brownish color was observed. The tissues were then rehydrated, immersed in xylol, covered with object glass, and examined under a microscope (Olympus Type CX-31, Tokyo, Japan). The number of apoptotic cells was counted by two independent observers.

4.4. Data Analysis

Data analysis was conducted using SPSS 18.0 for Windows (SPSS Inc. Chicago, IL, USA). The statistical tests used were one-way ANOVA for comparison between groups in each experimental design and the *t*-test for comparison between experimental designs. $p < 0.05$ was considered statistically significant.

5. Conclusions

Topical curcumin can act as a photoprotective agent by preventing cell apoptosis in UVB-induced mice. In terms of daily application and increasing concentration, 100 µM curcumin treatment was associated with the smallest number of histopathologically observed apoptotic cells after a single UVB exposure. However, at low concentrations, a single application of curcumin was more beneficial than daily application. Further studies, especially using concentrations lower than 0.1 µM, are warranted to prove this hypothesis.

Author Contributions: Conceptualization, K.D., N.M. and I.Y.; methodology, U.A.M., I.J.P. and M.F.; software, I.J.P. and M.F.; validation, K.D., M.N. and U.A.M.; formal analysis, I.J.P. and M.F.; investigation, K.D.; resources, K.D.; data curation, I.J.P. and M.F.; writing—original draft preparation, K.D.; writing—review and editing, K.D., U.A.M. and M.F.; visualization, I.J.P., U.A.M. and M.F.; supervision, M.N.M. and I.Y.; project administration, K.D. and M.F.; funding acquisition, K.D. and I.J.P. All authors have read and agreed to the published version of the manuscript.

Funding: This study is partly supported by a grant from the research institution of Hasanuddin University, Makassar, Indonesia (grant number 641/UN41/KEP/2019).

Institutional Review Board Statement: This study obtained ethics approval from the local ethics committee board (UH07100058).

Informed Consent Statement: Not applicable.

Data Availability Statement: The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Acknowledgments: We acknowledge Jonathan Kurnia Wijaya for his technical assistance.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of the compounds are available from the authors.

References

1. Tuorkey, M.J. Curcumin a Potent Cancer Preventive Agent: Mechanisms of Cancer Cell Killing. *Interv. Med. Appl. Sci.* **2014**, *6*, 139–146. [[CrossRef](#)] [[PubMed](#)]
2. Mirzaei, H.; Shakeri, A.; Rashidi, B.; Jalili, A.; Banikazemi, Z.; Sahebkar, A. Phytosomal Curcumin: A Review of Pharmacokinetic, Experimental and Clinical Studies. *Biomed. Pharmacother.* **2017**, *85*, 102–112. [[CrossRef](#)] [[PubMed](#)]
3. Abbaspour, H.; Afshar, A.S.; Safipour Afshar, A. Anti-Proliferative and Cytotoxic Effects of Curcumin in MCF-7 Human Breast Cancer Cells. *J. Chem. Health Risks* **2018**, *8*, 127–133. [[CrossRef](#)]
4. Elfahmi, N.V. *Phytochemical and Biosynthetic Studies of Lignans, with a Focus on Indonesian Medicinal Plants*; University of Groningen: Groningen, The Netherlands, 2006.
5. Kuno, T.; Tsukamoto, T.; Hara, A.; Tanaka, T. Cancer Chemoprevention through the Induction of Apoptosis by Natural Compounds. *J. Biophys. Chem.* **2012**, *3*, 156–173. [[CrossRef](#)]
6. Deng, H.; Wan, M.; Li, H.; Chen, Q.; Li, R.; Liang, B.; Zhu, H. Curcumin Protection against Ultraviolet-Induced Photo-Damage in Hacat Cells by Regulating Nuclear Factor Erythroid 2-Related Factor 2. *Bioengineered* **2021**, *12*, 9993–10006. [[CrossRef](#)]
7. Salucci, S.; Burattini, S.; Battistelli, M.; Baldassarri, V.; Maltarello, M.C.; Falciari, E. Ultraviolet B (UVB) Irradiation-Induced Apoptosis in Various Cell Lineages In Vitro. *Int. J. Mol. Sci.* **2012**, *14*, 532–546. [[CrossRef](#)]
8. Adusumilli, N.C.; Mordorski, B.; Nosanchuk, J.; Friedman, J.M.; Friedman, A.J. Curcumin Nanoparticles as a Photoprotective Adjuvant. *Exp. Dermatol.* **2021**, *30*, 705–709. [[CrossRef](#)]
9. Joshi, P.; Joshi, S.; Semwal, D.; Bisht, A.; Paliwal, S.; Dwivedi, J.; Sharma, S. Curcumin: An Insight into Molecular Pathways Involved in Anticancer Activity. *Mini Rev. Med. Chem.* **2021**, *21*, 2420–2457. [[CrossRef](#)]
10. Singh, R.; Letai, A.; Sarosiek, K. Regulation of Apoptosis in Health and Disease: The Balancing Act of BCL-2 Family Proteins. *Nat. Rev. Mol. Cell Biol.* **2019**, *20*, 175–193. [[CrossRef](#)]
11. Ghobrial, I.M.; Witzig, T.E.; Adjei, A.A. Targeting Apoptosis Pathways in Cancer Therapy. *CA Cancer J. Clin.* **2005**, *55*, 178–194. [[CrossRef](#)]
12. Tsai, K.-D.; Lin, J.-C.; Yang, S.-M.; Tseng, M.-J.; Hsu, J.-D.; Lee, Y.-J.; Cherng, J.-M. Curcumin Protects against UVB-Induced Skin Cancers in SKH-1 Hairless Mouse: Analysis of Early Molecular Markers in Carcinogenesis. *Evid. Based Complement. Altern. Med.* **2012**, *2012*, 593952. [[CrossRef](#)] [[PubMed](#)]
13. Ben Yehuda Greenwald, M.; Frušić-Zlotkin, M.; Soroka, Y.; Ben Sasson, S.; Bitton, R.; Bianco-Peled, H.; Kohen, R. Curcumin Protects Skin against UVB-Induced Cytotoxicity via the Keap1-Nrf2 Pathway: The Use of a Microemulsion Delivery System. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 5205471. [[CrossRef](#)] [[PubMed](#)]
14. Rainey, N.E.; Moustapha, A.; Petit, P.X. Curcumin, a Multifaceted Hormetic Agent, Mediates an Intricate Crosstalk between Mitochondrial Turnover, Autophagy, and Apoptosis. *Oxidative Med. Cell. Longev.* **2020**, *2020*, 3656419. [[CrossRef](#)]
15. Laszló, I.-P.; Laszló, M.R.; Popescu, T.; Toma, V.; Ion, R.M.; Moldovan, R.; Filip, G.A.; Cainap, C.; Clichici, S.; Muresan, A. The Comparative Effects of Resveratrol and Curcumin in Combination with Photodynamic Therapy. *Med. Pharm. Rep.* **2022**, *95*, 165–178. [[CrossRef](#)]
16. Li, H.; Gao, A.; Jiang, N.; Liu, Q.; Liang, B.; Li, R.; Zhang, E.; Li, Z.; Zhu, H. Protective Effect of Curcumin Against Acute Ultraviolet B Irradiation-Induced Photo-Damage. *Photochem. Photobiol.* **2016**, *92*, 808–815. [[CrossRef](#)]
17. Chan, W.-H.; Wu, C.-C.; Yu, J.-S. Curcumin Inhibits UV Irradiation-Induced Oxidative Stress and Apoptotic Biochemical Changes in Human Epidermoid Carcinoma A431 Cells. *J. Cell. Biochem.* **2003**, *90*, 327–338. [[CrossRef](#)] [[PubMed](#)]
18. Lou, S.; Wang, Y.; Yu, Z.; Guan, K.; Kan, Q. Curcumin Induces Apoptosis and Inhibits Proliferation in Infantile Hemangioma Endothelial Cells via Downregulation of MCL-1 and HIF-1 α . *Medicine* **2018**, *97*, e9562. [[CrossRef](#)]
19. Barbalho, S.M.; de Sousa Gonzaga, H.F.; de Souza, G.A.; de Alvares Goulart, R.; de Sousa Gonzaga, M.L.; de Alvarez Rezende, B. Dermatological Effects of Curcuma Species: A Systematic Review. *Clin. Exp. Dermatol.* **2021**, *46*, 825–833. [[CrossRef](#)]
20. Ali, R.E.; Rattan, S.I.S. Curcumin's Biphasic Hormetic Response on Proteasome Activity and Heat-Shock Protein Synthesis in Human Keratinocytes. *Ann. N. Y. Acad. Sci.* **2006**, *1067*, 394–399. [[CrossRef](#)]
21. McPherson, P.A.C.; McKenna, N.; Clare, D. Biphasic Effect of Curcuminoids on Oxidation of Postprandial Chylomicrons. *J. Med. Food* **2021**, *24*, 1340–1343. [[CrossRef](#)]
22. Zhou, H.-Y.; Sun, Y.-Y.; Chang, P.; Huang, H.-C. Curcumin Inhibits Cell Damage and Apoptosis Caused by Thapsigargin-Induced Endoplasmic Reticulum Stress Involving the Recovery of Mitochondrial Function Mediated by Mitofusin-2. *Neurotox. Res.* **2022**, *40*, 449–460. [[CrossRef](#)] [[PubMed](#)]
23. Saghir, S.A.M.; Alharbi, S.A.; Al-Garadi, M.A.; Al-Gabri, N.; Rady, H.Y.; Olama, N.K.; Abdulghani, M.A.M.; Al Hroob, A.M.; Almainan, A.A.; Bin-Jumah, M.; et al. Curcumin Prevents Cyclophosphamide-Induced Lung Injury in Rats by Suppressing Oxidative Stress and Apoptosis. *Processes* **2020**, *8*, 127. [[CrossRef](#)]
24. Park, K.; Lee, J.-H. Photosensitizer Effect of Curcumin on UVB-Irradiated HaCaT Cells through Activation of Caspase Pathways. *Oncol. Rep.* **2007**, *17*, 537–540. [[CrossRef](#)] [[PubMed](#)]
25. Ray, P.D.; Huang, B.-W.; Tsuji, Y. Reactive Oxygen Species (ROS) Homeostasis and Redox Regulation in Cellular Signaling. *Cell. Signal.* **2012**, *24*, 981–990. [[CrossRef](#)]
26. Liu, X.; Zhang, R.; Shi, H.; Li, X.; Li, Y.; Taha, A.; Xu, C. Protective Effect of Curcumin against Ultraviolet A Irradiation-induced Photoaging in Human Dermal Fibroblasts. *Mol. Med. Rep.* **2018**, *17*, 7227–7237. [[CrossRef](#)] [[PubMed](#)]
27. Pari, L.; Tewas, D.; Eckel, J. Role of Curcumin in Health and Disease. *Arch. Physiol. Biochem.* **2008**, *114*, 127–149. [[CrossRef](#)]

28. Kong, Y.; Li, M.; Guo, G.; Yu, L.; Sun, L.; Yin, Z.; Li, R.; Chen, X.; Wang, G. Effects of Dietary Curcumin Inhibit Deltamethrin-Induced Oxidative Stress, Inflammation and Cell Apoptosis in *Channa Argus* via Nrf2 and NF-KB Signaling Pathways. *Aquaculture* **2021**, *540*, 736744. [[CrossRef](#)]
29. Hybertson, B.M.; Gao, B.; Bose, S.K.; McCord, J.M. Oxidative Stress in Health and Disease: The Therapeutic Potential of Nrf2 Activation. *Mol. Asp. Med.* **2011**, *32*, 234–246. [[CrossRef](#)]
30. Li, M.; Kong, Y.; Wu, X.; Guo, G.; Sun, L.; Lai, Y.; Zhang, J.; Niu, X.; Wang, G. Effects of Dietary Curcumin on Growth Performance, Lipopolysaccharide-Induced Immune Responses, Oxidative Stress and Cell Apoptosis in Snakehead Fish (*Channa Argus*). *Aquac. Rep.* **2022**, *22*, 100981. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

Single or Daily Application of Topical Curcumin Prevents Ultraviolet B-Induced Apoptosis in Mice

ORIGINALITY REPORT

19%

SIMILARITY INDEX

14%

INTERNET SOURCES

15%

PUBLICATIONS

6%

STUDENT PAPERS

PRIMARY SOURCES

1	www.dovepress.com Internet Source	2%
2	pubmed.ncbi.nlm.nih.gov Internet Source	1%
3	www2.mdpi.com Internet Source	1%
4	www.turmeric-curcumin.com Internet Source	1%
5	jbiomedsci.biomedcentral.com Internet Source	1%
6	Jaw-Ming Cherng. "Diallyl sulfide protects against ultraviolet B-induced skin cancers in SKH-1 hairless mouse: analysis of early molecular events in carcinogenesis : Prevention of photocarcinogenesis by diallyl sulfide", Photodermatology Photoimmunology & Photomedicine, 06/2011 Publication	1%

7	Internet Source	1 %
8	Submitted to Universidad Anahuac México Sur Student Paper	1 %
9	Mariella Nieddu, Federica Pollastro, Paola Caria, Stefano Salamone, Antonella Rosa. "Xanthomicrol Activity in Cancer HeLa Cells: Comparison with Other Natural Methoxylated Flavones", Molecules, 2023 Publication	1 %
10	www.tdx.cat Internet Source	1 %
11	Vandita, Kakkar, Bhushan Shashi, Kumar Guru Santosh, and Kaur Indu Pal. "Enhanced Apoptotic Effect of Curcumin Loaded Solid Lipid Nanoparticles", Molecular Pharmaceutics, 2012. Publication	1 %
12	journals.plos.org Internet Source	1 %
13	www.uniklinikum-dresden.de Internet Source	<1 %
14	diagnosticpathology.biomedcentral.com Internet Source	<1 %
15	spandidos-publications.com	

<1 %

16

triggered.edina.clockss.org

Internet Source

<1 %

17

Chenghui Pan, Yonggang Yan, Dayun Zhao. "The Fate and Intermediary Metabolism of Soyasapogenol in the Rat", *Molecules*, 2022

Publication

<1 %

18

Ekaterina Proshkina, Mikhail Shaposhnikov, Alexey Moskalev. "Genome-Protecting Compounds as Potential Geroprotectors", *International Journal of Molecular Sciences*, 2020

Publication

<1 %

19

mdpi.com

Internet Source

<1 %

20

Koh, S.H.. "Phosphatidylinositol-3 Kinase/Akt and GSK-3 Mediated Cytoprotective Effect of Epigallocatechin Gallate on Oxidative Stress-Injured Neuronal-Differentiated N18D3 Cells", *Neurotoxicology*, 200409

Publication

<1 %

21

V. Cekarini, M. Cuccioloni, M. Mozzicafreddo, L. Bonfili, M. Angeletti, A. M. Eleuteri. "Targeting Proteasomes with Naturally Occurring Compounds in Cancer Treatment", *Current Cancer Drug Targets*, 2011

<1 %

22 Weiguang Zou, Yaobin Ma, Chunxiang Ai, Wenchao Yu, Xiaolong Gao, Shengtai Liu, Xuan Luo, Weiwei You, Caihuan Ke. "Dietary curcumin influence on growth, antioxidant status, immunity, gut flora and resistance to *Vibrio harveyi* AP37 in *Haliotis discus hannai*", *Aquaculture Reports*, 2022
Publication

23 res.mdpi.com
Internet Source

24 www.wjgnet.com
Internet Source

25 Layasadat Khorsandi, Mehri Mirhoseini, Masoomah Mohamadpour, Mahmoud Orazizadeh, Soheila Khaghani. "Effect of curcumin on dexamethasone-induced testicular toxicity in mice", *Pharmaceutical Biology*, 2012
Publication

26 Milisav, Irina, Borut Poljsak, and Dušan Šuput. "Adaptive Response, Evidence of Cross-Resistance and Its Potential Clinical Use", *International Journal of Molecular Sciences*, 2012.
Publication

27 s-space.snu.ac.kr
Internet Source

<1 %

28

wjgnet.com

Internet Source

<1 %

29

"FREE COMMUNICATIONS POSTER COMMUNICATIONS", Nephrology, 6/2005

Publication

<1 %

30

Heba Mohammed Fadhil, Mohammed Najm Abdullah, Mohammed Issam Younis. "TWGH: A Tripartite Whale-Gray Wolf-Harmony Algorithm to Minimize Combinatorial Test Suite Problem", Electronics, 2022

Publication

<1 %

31

Laura Marinela Ailioaie, Gerhard Litscher. "Curcumin and Photobiomodulation in Chronic Viral Hepatitis and Hepatocellular Carcinoma", International Journal of Molecular Sciences, 2020

Publication

<1 %

32

Min Sung Kim, Yong Tae Ahn, Chul Won Lee, Hyungwoo Kim, Won Gun An. "Astaxanthin Modulates Apoptotic Molecules to Induce Death of SKBR3 Breast Cancer Cells", Marine Drugs, 2020

Publication

<1 %

33

Wen-Hsiung Chan, Hsin-Jung Wu. "Anti-apoptotic effects of curcumin on

<1 %

photosensitized human epidermal carcinoma A431 cells", Journal of Cellular Biochemistry, 2004

Publication

34

eresources.uin-malang.ac.id

Internet Source

<1 %

35

ijmhs.biomedcentral.com

Internet Source

<1 %

36

www.tandfonline.com

Internet Source

<1 %

37

Ben-Yehuda Greenwald, Maya, Shmuel Ben-Sasson, Havazelet Bianco-Peled, and Ron Kohen. "Skin Redox Balance Maintenance: The Need for an Nrf2-Activator Delivery System", *Cosmetics*, 2016.

Publication

<1 %

38

Suhui Lou, Yanfang Wang, Zujiang Yu, Kelei Guan, Quancheng Kan. "Curcumin induces apoptosis and inhibits proliferation in infantile hemangioma endothelial cells via downregulation of MCL-1 and HIF-1 α ", *Medicine*, 2018

Publication

<1 %

39

Weilan Wan, Zhiqi Hou, Qiuying Qiu. "Postoperative analgesic effect of dexmedetomidine combined with TPVB

<1 %

applied to open gastrectomy for gastric cancer", Research Square Platform LLC, 2022

Publication

40

Hicham Oualla, Rachid Fateh, Anouar Darif, Said Safi, Mathieu Pouliquen, Miloud Frikel. "Channel Identification Based on Cumulants, Binary Measurements, and Kernels", Systems, 2021

Publication

<1 %

Exclude quotes Off

Exclude matches Off

Exclude bibliography On